



PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of

Xiao and Gedrich

Serial No. 09/879,792

Filing Date: June 13, 2001

)
) Group Art Unit: 1652
)
) Examiner: D. Ramirez
)
) Docket No. 002973.00035
)

RECEIVED
JUN 11 2003
TECH CENTER 1600/2900

For: **REGULATION OF HUMAN TRANSMEMBRANE SERINE PROTEASE**

DECLARATION UNDER RULE 131

We, Yonghong Xiao and Richard Gedrich, hereby declare:

1. We are the named inventors of the subject matter claimed in the application referenced above.
2. Prior to March 19, 2001, we conceived of and reduced to practice molecules comprising the polynucleotide sequence disclosed as SEQ ID NO:11 and the amino acid sequence disclosed as SEQ ID NO:12 in the above-referenced application.
3. The nucleotide sequence of a contig identified as SEQ ID NO:11 in serial number 60/211,224 was extended by amplification and alignments with mouse EST and human genomic sequences. The nucleotide sequence of an extended contig containing a portion of the sequence of SEQ ID NO:11 in serial number 09/879,792 was identified. The nucleotide sequence of the extended contig is shown on the attached copy of page 3 of laboratory notebook RB55202 and identified as "147.33 contig" (Exhibit A).
4. The Basic Local Alignment Search Tool (BLAST) algorithm was used to extend the 5' end sequence of the 147.33 contig. The BLAST algorithm aligned human expressed sequence tags (ESTs) having sequence similarity with the 5' sequence of the 147.33 contig. The results of the BLAST searches were recorded on pages 20-23 of laboratory notebook RB55202. Copies are provided as Exhibit B.
5. An EST, BE280394, was found to overlap the 5' sequence of the 147.33 contig. The BE280394 EST sequence was recorded on page 24 of laboratory notebook RB55202. A copy

of page 24 is provided as Exhibit C. The alignment of EST BE280394 with the 5' end of the contig is shown in second column of page 20, the first column of page 21, and the first lines of the second column of page 21 (Exhibit B).

6. To confirm that the 147.33 contig extended at its 5' end with the sequence of EST BE280394 included the sequence of a full-length gene, the extended 147.33 contig was translated by computer into an amino acid sequence. The translation of the extended contig in all possible open reading frames associated with the extended sequence was recorded on pages 27 and 28 of laboratory notebook RB55202. Copies are provided as Exhibit D. The extended contig was confirmed to contain a full-length gene because its translation into an amino acid sequence identified a start (methionine) residue at its amino terminal end and the start codon is in frame with a predicted amino acid sequence encoded by the 147.33 contig. The translated sequence identified as the one encoding the protein shown in SEQ ID NO:12 is underlined and begins "MERDSH." The hand-written notes at the top of page 29 of laboratory notebook RB55202 identify the extended contig sequence as the sequence of the "147" full-length gene. A copy of page 29 is included in Exhibit D.

7. Primers capable of amplifying the 147 full-length open reading frame were ordered based on the nucleotide sequence that encodes the translated protein. The oligonucleotide sequences ordered were recorded on page 30 of laboratory notebook RB55202, a copy of which is provided as Exhibit E.

8. Human placenta cDNA was amplified using these primers to obtain the molecule in human genomic DNA that contains the 147 full-length open reading frame. The primers used to perform the amplification and the conditions of the amplification reaction were recorded on page 40 of laboratory notebook RB55202. A copy of page 40 is provided as Exhibit F.

9. The amplification product obtained from this reaction was the predicted size for the full-length open reading frame. An agarose gel showing the amplification product obtained using the primers is shown in a photograph on page 41 of laboratory notebook RB55202. A copy of page 41 is provided as Exhibit G.

10. The amplification product was extracted from the gel using the "QIAquick Gel Extraction Kit Protocol" and cloned into a vector sold by Invitrogen and referred to the "TA vector." The protocol used to clone the amplification product (full-length open reading frame)

into the vector was recorded on pages 41 and 42 of lab notebook RB55202. Copies of pages 41 and 42 are provided as Exhibits G and H, respectively.

11. The vector containing the amplification product was transformed into *E. coli* TOP10 cells, as described on page 42 of laboratory notebook RB55202 (Exhibit H).

12. The vector was extracted from the transformed *E. coli* and digested with restriction enzyme *EcoRI* to confirm the presence of the amplification product in the vector. The protocol "QIAprep Spin Miniprep Kit Protocol," which was used to obtain the vector DNA from *E. coli*, was recorded on page 1 of laboratory notebook RB55846. A copy of page 1 is included in Exhibit I. The conditions under which the vector DNA was digested and a picture of the agarose gel showing the results of restriction enzyme digestion are provided on page 7 of laboratory notebook RB55846. A copy of this page is included in Exhibit I. The agarose gel on the laboratory notebook page confirms the presence of the 147 full-length open reading frame in each vector but for one.

13. Vectors containing the amplification product were sequenced. The composition of the tubes containing the vector and primers used in the sequencing reaction were recorded on page 8 of laboratory notebook RB55846. A copy of page 8 is provided as Exhibit J.

14. The nucleotide sequence obtained for amplification product cloned into the vector was recorded on pages 15-17 of laboratory notebook RB55846. Copies of pages 15-17 are provided as Exhibit K. The recorded nucleotide sequence is identical to the sequence identified as SEQ ID NO:11 in application serial number 09/879,792.

15. The nucleotide sequence of the amplification product was translated into an amino acid sequence. This translated amino acid sequence was recorded on pages 17-18 of laboratory notebook RB55846. Copies of pages 17-18 are provided as Exhibit L. The recorded amino acid sequence is identical to the sequence identified as SEQ ID NO:12 in application serial number 09/879,792.


16. The dates on each of the attached Exhibits have been redacted. All of the work described in paragraphs 3 through 15 was performed in the United States and completed before March 19, 2001.

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these were made with the knowledge that false statements made willfully are punishable by fine, imprisonment, or both a fine and imprisonment under Section 1001 of Title 18 of the United States; and further that false statements made willfully may jeopardize the validity of any patent issuing on an application in which the false statements were made.

Date

Yonghong Xiao

6/5/03
Date

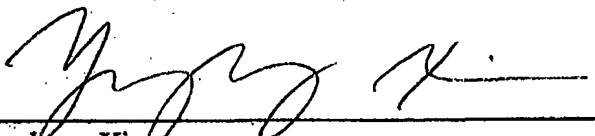


Richard Gedrich

We hereby declare that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these were made with the knowledge that false statements made willfully are punishable by fine, imprisonment, or both a fine and imprisonment under Section 1001 of Title 18 of the United States; and further that false statements made willfully may jeopardize the validity of any patent issuing on an application in which the false statements were made.

6/9/03

Date


Yonghong Xiao~~6/9/03~~

Date


Richard Gedrich

BAYER CORPORATION

Note Book 3
RB55202

SUBJECT

147.33.contig2 Sequen (Nucleotide)

EXHIBIT

A

147.33.contig2
(Stream)

1 GAATTCGCTT TTGCTTTTCA AGTCATCTTC CAGCAAGAAC
41 ACCTTCAGCT GGAGCATCTC CAGCCGAGGT ATCTCCAGCT
61 GGGACACCTC CAGGCGCGGC ATCTCCAGCT CAGGATCTTC
121 CAGCCGAGGT ATCTCCAGCT GGGACACCTC CAGGCGCGGC
161 ATCTCCAGCT CAGGATCTTC CAGCTGGTAC AGCTCCAGCT
201 CAGGATCTTC CAGGCGCGGC ATCTCCAGCT CAGGATCTTC
241 CAGCTCCAGG ATCTCCAGCT CAGGATCTTC ATCTCCAGGT
281 CTGATCCGCT AGGTCATCAT CCGCCAGGTC AGCTCCAGGT
321 ACAAGCTCCG CAGCCGAGGT GTACCTCTCT AGAGCAACAC
361 CAGTGGGGGC TTGACCCATC GATCATCTTC CTGCGAGGTC
401 AGCAGCCGCA ACCGAGCCCA CAGGCGAGG CCGAGGTACC
441 AGCTTCGCTC AGTTCCAGCT CCGGAGGAGG CAGAGCCAGC
481 TACCCGCTAT TCCGCTGGGT CTCTCTCTCA TTGCGCTGCT
521 GCTTCTGCTC ATCATCTCTC TCGATCTCTC CAGGCGCGGC
561 ACAGGGGTCA GGTCAAGGCA CCGAGGAGG AGCTTCGCTC
601 AGCAGCTGCT TCCGCTGGGT CTCTCTCTCA TTGCGCTGCT
641 GAAGAGTCA CAGCTCCAGT CCGTGGAGGT TGACTCGGAC
681 AAGTCTCTGC TCAAACTCTA CTCTCTCTCA TCCCATCTCT
721 GGTCTCTCAT CTCTAGCAGC AACTGGAATG AGCTCTCTCT
761 ACAGAGGAGC TCCGAGGAGC TGGGTTTCCA GATCTCTGAG
801 CCGACACCGG AGTTTGGCCA CAGGGAATTT CCGACAGGCT
841 TCTCAATCTT GAGATGAGC TCCAGCATCT ACCAAGGCT
881 CCGAGGCTCT GAATGCGCTT CCGAGCGCTA TATCTCTCTC
921 CAGTCTCTCC ACTGCGGACT CAGGCGATAG ACCGCGCGCA
961 TCGTGGGAGG GCGGCTGCGC TCGCATAGCA AGTGGCTCTC
1001 GCAAGTGAAT CTGCACTTTC GCACCAACCA CATCTCTGCA
1041 GGCAGCCTCA TTGACCCCCA GTGGGTGCTC ACTGCGCGCC
1081 ACTGCTCTCT CCGTACCCCG GAGAGGTTCC TCGAGCGCTC
1121 CAGGCTCTAC GCGGCGAGCA GCAACCTCCA CAGTTGCTCT
1161 CAGGCGAGCT CCAATGCGCA GATCATCTTC AACAGCAAT
1201 ACACCGATGA GGAAGGAGC TATGACATCG CCGTCACTGC
1241 GGTGCTCAAG CCGCTGAGCC TGTCTGCTCA CATCCACCTC
1281 GGTGCTCTCC CCAATGAGTC ACAGAGCTTT AGCTCAATG
1321 AGACCTCTCT GATCAGAGC TTGCGGAGG CCGAGGAGG
1361 AGATGACAG ACATGCGCTT TCGTCCGGA GGTGAGGTC
1401 AATCTCATCG ACTTCAAGAA ATGCAATGAC TACTTGGTCT
1441 ATGACAGTGA CTTTACGCGA AGGATCATCT GTGCTGGGCA
1481 CTTTCTGGGG GGCAGAGACT CCGTCCAGGG AGACAGCGCG
1521 GCGGCTCTTC TGTGAGCA GAACAACCG TGGTACCTGC
1561 CAGGTGTGAC CAGCTGGGCG CAGCTCTGTG GCGAGAGAA
1601 CAACCTGCTT CTGTACACCA AAGTGACAGA ACTTCTTCTC
1641 TCGATTTACA CCAACATGGA GAGGAGGTC CCAATCAGAA
1681 AATCTTAACC AGCTGCGCTG CTCTCTGCA CAGCAGCGCC
1721 TCTCTGACT AAGGGCGAAT TC

LifeTools Reverse FASTA Sequences



Retrieve
FASTA Sequences

FASTA Format FASTA Format FASTA Format FASTA Format FASTA Format FASTA Format FASTA Format

Project ID: 000159147 Project Name: TM Series Proteome
Created By: Gedrich, Richard Created By: Gedrich, Richard
Modified By: Gedrich, Richard Modified By: Gedrich, Richard
Status: OPEN Current User: Gedrich, Richard (WHITE)
Today's Date: 06/06/00 Run Seqs: 26

Reverse(147.33.42.75contig2.1)
GAATTCGCTT TTGCTTTTCA AGTCATCTTC CAGCAAGAAC
ACCTTCAGCT GGAGCATCTC CAGCCGAGGT ATCTCCAGCT
GGGACACCTC CAGGCGCGGC ATCTCCAGCT CAGGATCTTC
CAGCCGAGGT ATCTCCAGCT GGGACACCTC CAGGCGCGGC
ATCTCCAGCT CAGGATCTTC CAGCTGGTAC AGCTCCAGCT
CAGGATCTTC CAGGCGCGGC ATCTCCAGCT CAGGATCTTC
CAGCTCCAGG ATCTCCAGCT CAGGATCTTC ATCTCCAGGT
CTGATCCGCT AGGTCATCAT CCGCCAGGTC AGCTCCAGGT
ACAAGCTCCG CAGCCGAGGT GTACCTCTCT AGAGCAACAC
CAGTGGGGGC TTGACCCATC GATCATCTTC CTGCGAGGTC
AGCAGCCGCA ACCGAGCCCA CAGGCGAGG CCGAGGTACC
AGCTTCGCTC AGTTCCAGCT CCGGAGGAGG CAGAGCCAGC
TACCCGCTAT TCCGCTGGGT CTCTCTCTCA TTGCGCTGCT
GCTTCTGCTC ATCATCTCTC TCGATCTCTC CAGGCGCGGC
ACAGGGGTCA GGTCAAGGCA CCGAGGAGG AGCTTCGCTC
AGCAGCTGCT TCCGCTGGGT CTCTCTCTCA TTGCGCTGCT
GAAGAGTCA CAGCTCCAGT CCGTGGAGGT TGACTCGGAC
AAGTCTCTGC TCAAACTCTA CTCTCTCTCA TCCCATCTCT
GGTCTCTCAT CTCTAGCAGC AACTGGAATG AGCTCTCTCT
ACAGAGGAGC TCCGAGGAGC TGGGTTTCCA GATCTCTGAG
CCGACACCGG AGTTTGGCCA CAGGGAATTT CCGACAGGCT
TCTCAATCTT GAGATGAGC TCCAGCATCT ACCAAGGCT
CCGAGGCTCT GAATGCGCTT CCGAGCGCTA TATCTCTCTC
CAGTCTCTCC ACTGCGGACT CAGGCGATAG ACCGCGCGCA
TCGTGGGAGG GCGGCTGCGC TCGCATAGCA AGTGGCTCTC
GCAAGTGAAT CTGCACTTTC GCACCAACCA CATCTCTGCA
GGCAGCCTCA TTGACCCCCA GTGGGTGCTC ACTGCGCGCC
ACTGCTCTCT CCGTACCCCG GAGAGGTTCC TCGAGCGCTC
CAGGCTCTAC GCGGCGAGCA GCAACCTCCA CAGTTGCTCT
CAGGCGAGCT CCAATGCGCA GATCATCTTC AACAGCAAT
ACACCGATGA GGAAGGAGC TATGACATCG CCGTCACTGC
GGTGTCAAG CCGCTGAGCC TGTCTGCTCA CATCCACCTC
GGTGTCTCTC CCAATGAGTC ACAGAGCTTT AGCTCAATG
AGACCTCTCT GATCAGAGC TTGCGGAGG CCGAGGAGG
AGATGACAG ACATGCGCTT TCGTCCGGA GGTGAGGTC
AATCTCATCG ACTTCAAGAA ATGCAATGAC TACTTGGTCT
ATGACAGTGA CTTTACGCGA AGGATCATCT GTGCTGGGCA
CTTCTGGGG GGCAGAGACT CCGTCCAGGG AGACAGCGCG
GCGGCTCTTC TGTGAGCA GAACAACCG TGGTACCTGC
CAGGTGTGAC CAGCTGGGCG CAGCTCTGTG GCGAGAGAA
CAACCTGCTT CTGTACACCA AAGTGACAGA ACTTCTTCTC
TCGATTTACA CCAACATGGA GAGGAGGTC CCAATCAGAA
AATCTTAACC AGCTGCGCTG CTCTCTGCA CAGCAGCGCC
TCTCTGACT AAGGGCGAAT TC

Requested by: Gedrich, Richard on Tue

INCYTE PHARMACEUTICALS, INC.

SIGNED BY

[Signature]

DATE

WITNESSED AND UNDERST

D BY

[Signature]

DATE

CROSS REFERENCES:

BAYER CORPORATION

RB55202 21

SUBJECT

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27

EXHIBIT

D

LifeTools
In Sport, as in War

[illegible]

Page 6-3	Page 1	100%	100%	100%	100%	100%
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Protein (g/100g)	Carbohydrate (g/100g)	Moisture (g/100g)	Energy (kJ/100g)	Energy (kcal/100g)	Energy (kJ/g)	Energy (kcal/g)	Energy (kJ/kg)	Energy (kcal/kg)
Protein (g/100g)	Carbohydrate (g/100g)	Moisture (g/100g)	Energy (kJ/100g)	Energy (kcal/100g)	Energy (kJ/g)	Energy (kcal/g)	Energy (kJ/kg)	Energy (kcal/kg)

COOLEST: 47, NE20004:2

Sequence: coolERT147:RE280394:2

	300	350	360	350	360

+1 K T E Z V T D A V L A - G I D W D O R L
 +2 R L E N - O T L C - R B V L T O T N V S

28

29

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CRNEE DEPENDENCE.

REST202 29

Conclusions (continued)

* MERDSH appears to be what I had located

The AT6 here looks to be a great Kozak consensus

ACCATGG.

This is it.

= ORDER PRIMERS

LBRI #147 amino acid Sequence

NERDSHGNASPARTPSAGASPAQVSPAGTPPGRASPAQASPAQASPAQTPPGRASPAQASPAQTPPGRASPG
RASPAQASPAQASPALASLSRSSGRSSASASVTTSPTRVYLVRATPVGAVPIRSSPARSAPATRATRES
PGTSLPKFTWREGQKQLPLIGCVLLLIALLVLSLIILFQFWQGHGTGIRYKEQRESCPKHAVRCDGVVDCKLKS
DELQCVRFDFWDKSLKKIYSGSSHQWLPCSSNNWDSYSEKTCQQLGFESAHRTEVAHRDFANSFSILRYNS
TIQESLRHSECPQRSYISLQCSHQLCRAMTGRIVGGALASDSKNWPQVSLHFGTTHICGGLIDAQWVLTA
HCFFVTREKVLGKWIYAGTCSNLHQLPEAAETIINSNYTDEEDDYIALMRYSKPLTSAHIHPACLPM
HGQTFSLNETCWTGFGKTRETDDKTSPLFREVQVNLIDFKKCNDDLYVDYSMLTPRMMCAGDLRGGRDSCQG
DSGGPLVCEQNNRWYLAGVTSWGAGCGQRNKGVYTKVTEVLPWIYSKMESEVFRKS

Start is an NcoI site.

My predicted sequence diverges from the 5' end of BE280394 and a consensus splice acceptor (CAG).

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CR SS REFERENCES:

BAYER CORPORATION

RB55202

SUBJECT

Oligo Order

EXHIBIT

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LBRI Oligos

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5

CoolEST 147- oligos to MY predicted genomic sequence

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7

RGL0054

Cloning primer 23-mer to BE280394 sequence

8

5' CTC AGA GAC CAT GGA GAG GGA CA 3'

9

10

RGL0055Sense Sequencing Primer 21-mer; bases 331-351 of
147.33.49.28contig2

11

12

5' CAA CCA GAG TGT ACC TTG TTA 3'

13

14

RGL0056antiense Sequencing Primer 20-mer; bases 441-460 of
147.33.49.28contig2

15

16

5' CAG GTG AAC TTG GGC AGG CT 3'

17

18

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Orcl

oligos to clone "Full length 147" shown on

Page 29

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D BY

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DATE

CR SS REFERENCES:

BAYER CORPORATION

RB55202

SUBJECT PCR with RGL0054/0028

EXHIBIT

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Purpose: to test hypothesis for the new 5' ends

Received oligos

336178
R7181C08 (C08)
RGL0056
Rich Gedrich
284.06 ng (28 ng/100) OD 13.99
MW: 6180.0 u/gmole read 62.1
CAGGTGACTTGGGAGGCT

336178
R7181C08 (C08)
RGL0056
Rich Gedrich
456.23 ng (28 ng/100) OD 17.56
MW: 7732.8 u/gmole read 64.2
CTCAGGACCATGGGAGGACA

336178
R7181C07 (C07)
RGL0054
Rich Gedrich
286.11 ng (28 ng/100) OD 13.80
MW: 7732.8 u/gmole read 69.9
CTCAGGACCATGGGAGGACA

Resuspended RGL0054 to 200nm

dilute 250ul dH₂Odilute 1/24 to 50ul (25ul + 75ul dH₂O)

in 50ul PCR

ZLWS

do plasmid COMA -/+ RT

in Advantage GC kit (Clontech)

Rows

1) - RT

2) + RT

Row

3ul (3)

2ul plasmid COMA

30ul

1) Aliq + PM

10ul 5X Buffer

3ul

2) Add COMA

1ul Advantage GC polymerase

3ul

3) Cycle in PCR600

1ul 10mM dNTP

3ul

1ul RGL0054

3ul

94°C 1' → 54°C 15" → 68°C 35" } 22 cycles
1 cycle } 1 cycle of

1ul RGL0028

3ul

5ul GC mix

15ul

29ul dH₂O

82ul

50ul

48ul Row

SIGNED BY

R. L. W. M.

DATE

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Elizabeth C. Sullivan

DATE

CROSS REFERENCES:

BAYER CORPORATION

RB55202

41

SUBJECT



Run 5-plate on 1.2% Agarose gel (1X TAE)

M = 1 Kb Ladder (GIBCO)

The Run was made. A ~1.8 Kb Band was seen in the +RT Run only

Clone not PCR'd

Run Remnants of the sample on a 1.2% Agarose gel

Cut out the Fragment

QIAquick Gel Extraction Kit Protocol

using a microcentrifuge

This protocol is designed to extract and purify DNA of 10 bp to 10 kb from standard or low melt agarose gels in TAE or TBE buffer.

- Notes:
- **NEW** The yellow color of Buffer OG indicates a pH of 7.5.
 - Add ethanol (95-100%) to Buffer PE before use (see bottle label for volume).
 - Isopropanol (100%) will be required.
 - A heating block or water bath at 50°C is required.
 - All centrifugation steps are carried out at 10,000 x g (~13,000 rpm) in a conventional laboratory microcentrifuge.
 - 3M sodium acetate, pH 5.0, may be necessary.

1. Excise the DNA fragment from the agarose gel with a clean, sharp scalpel. Minimize the size of the gel slice by removing extra agarose.
2. Weigh the gel slice in a colorless tube. Add 3 volumes of Buffer OG to 1 volume of gel (100 mg ~ 100 µl).
For example, add 300 µl of Buffer OG to each 100 mg of gel for a 2% agarose gel. Add 1 volume of Buffer OG. The maximum amount of gel slice per QIAquick column is 400 mg; for gel slices >400 mg use more than one QIAquick column.
3. Incubate at 50°C for 10 min (or until the gel slice has completely dissolved). To help dissolve gel, mix by vortexing the tube every 2-3 min during the incubation.
IMPORTANT: Solubilize viscous completely. For >2% gels, increase incubation time.
4. After the gel slice has dissolved completely, check that the color of the mixture is yellow (similar to Buffer OG without dissolved agarose). If the color of the mixture is orange or violet, add 10 µl of 3M sodium acetate, pH 5.0, and mix. The color of the mixture will turn to yellow.
The adsorption of DNA to QIAquick membrane is efficient only at pH 7.5. Buffer OG contains a pH indicator which is yellow at pH 7.5 and orange or violet at higher pH. Following entry determination of the optimal pH for DNA binding.
5. Add 1 gel volume of isopropanol to the sample and mix.
For example, if the agarose gel slice is 100 mg, add 100 µl isopropanol. This step increases the yield of DNA fragments <500 bp and >4 kb. For DNA fragments between 500 bp and 4 kb, addition of isopropanol has no effect on yield. Do not centrifuge the sample at this stage.

6. Place a QIAquick spin column in a provided 2-ml collection tube.
7. To bind DNA, apply the sample to the QIAquick column, and centrifuge for 1 min.
The maximum volume of the column reservoir is 800 µl. For sample volumes of more than 800 µl, apply load and spin again.
8. Discard flow-through and place QIAquick column back in the same collection tube.
Collection tubes are reused to reduce plastic consumption.
9. (Optional): Add 0.5 ml of Buffer OG to QIAquick column and centrifuge for 1 min.
This step will remove all traces of agarose. It is only required for direct sequencing, in vitro transcription or microinjection.
10. To wash, add 0.75 ml of Buffer PE to QIAquick column and centrifuge for 1 min.
Notes: If the DNA will be used for soft-sphere applications, such as library construction and direct sequencing, let the column stand 2-5 min after addition of Buffer PE, before centrifuging.
11. Discard the flow-through and centrifuge the QIAquick column for an additional 1 min at 10,000 x g (~13,000 rpm).
IMPORTANT: Residual ethanol from Buffer PE will not be completely removed unless the flow-through is discarded before the additional centrifugation.
12. Place QIAquick column into a clean 1.5-ml microfuge tube.
13. To elute DNA, add 50 µl of Buffer EB (10 mM Tris-Cl, pH 8.5) or H₂O to the center of the QIAquick column and centrifuge for 1 min at maximum speed. Alternatively, for increased DNA concentration, add 30 µl elution buffer to the center of the QIAquick column, let stand for 1 min, and then centrifuge for 1 min.
IMPORTANT: Ensure that the elution buffer is dispensed directly onto the QIAquick membrane for complete elution of bound DNA. The average elution volume is 45 µl from 50 µl elution buffer volume, and 20 µl from 30 µl.
(Elution efficiency is dependent on pH. The maximum elution efficiency is achieved between pH 7.0 and 8.5. When using water, make sure that the pH value is within this range, and store DNA at -20°C as DNA may degrade in the absence of a buffering agent. The purified DNA can also be stored in TE (10 mM Tris-Cl, 1 mM EDTA, pH 8.0), but the DNA may exhibit subsequent enzymatic reactions.

SIGNED BY

WITNESSED AND UNDERST

BY

Elizabeth C. Sullivan

DATE

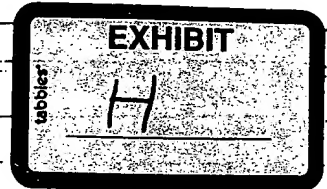
DATE

CROSS REFERENCES:

BAYER CORPORATION

RB55202

SUBJECT



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Eliel - 36ml dH₂O 147 0054/28 PER

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TA cloning kit (Invitrogen)

5

6

Ligate Frag into pCR

7

R₁

8

1ml vector

9

1ml 10x

10

4ml dH₂O

11

1ml Lysate

12

3ml Frag

13

1ml

14

16 °C o/n T₁ = 550pm

15

16

17

Transform TDP 10

18

2ml R₁ / ~~pre~~ take

19

- 1 hr 25'

20

- 42 °C 30"

21

- 1 hr 2'

22

Add 400 µl SOC

23

37 °C 40'

24

25

plus 100 µl on LB + Amp Plate + Bluejet

26

27

37 °C o/n T₁

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SIGNED BY

DATE

WITNESSED AND UNDERST

DATE

CR SS REFERENCE :

BAYER CORPORATION

RB5584L

7

SUBJECT LBRI #147

EXHIBIT

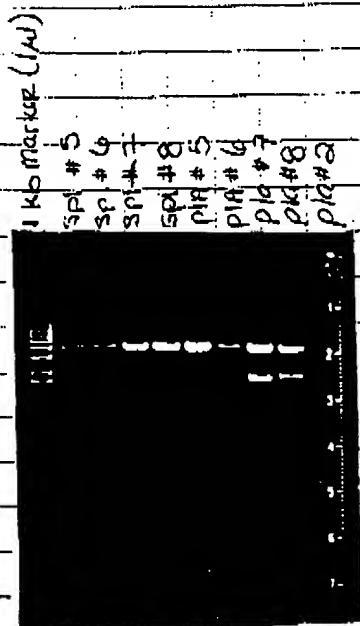
I

- ① miniprep of spl 5,6,7,8 : pla 5,6,7,8 : pla 2
 - * Followed same protocol from page 1 of this notebook.
 - * Eluted DNA in 50 μ l water

- ② Restriction enzyme digestion of pla 5,6,7,8, spl 5,6,7,8 : pla 2

Plasmid	3 μ l	Digest for 2 hrs @ 37°C
Eco RI	1 μ l	
Buffer 2	2 μ l	
H ₂ O	14 μ l	
	20 μ l	

- ③ RAN 1% Agarose Gel



* The miniprep from pla 2 gave negative results. Therefore, I did a new transformation using 1 μ l of a 1:200 dilution of my original pla 2 miniprep. Transformed in 30 \times of DH5 α max efficiency.

- 1 \times dilution + 30 \times cells
- incubate on ice for 5-10 minutes
- Add 100 \times SOC
- Plate 100 \times on Amp plates
- Incubate @ 37°C overnight

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DATE

DATE

BAYER CORPORATION

SUBJECT LARI #147

① Miniprep of #1475428 (LARI)
 Rich Gedrich gave me 4 clones from spleen
 and 4 clones from placenta

② I followed the following protocol using 1.5 ml culture

QIAprep Spin Miniprep Kit Protocol

using a microcentrifuge

This protocol is designed for purification of up to 20 µg of high-copy plasmid DNA from 1-5 ml overnight cultures of *E. coli* in LB [Luria-Bertani] medium. For purification of low-copy plasmids and cosmids, large plasmids (>10 kb), and DNA prepared using other methods, refer to the recommendations on page 31.

Please read Important Notes for QIAprep Procedures on pages 14-15 before starting.

Procedure

1. Resuspend pelleted bacterial cells in 250 µl of Buffer P1 and transfer to a microfuge tube.
 Ensure that RNase A has been added to Buffer P1. No cell clumps should be visible after resuspension of the pellet.
2. Add 250 µl of Buffer P2 and gently invert the tube 4-6 times to mix.
 Mix gently by inverting the tube. Do not vortex, as this will result in shearing of genomic DNA. If necessary, continue inverting the tube until the solution becomes viscous and slightly clear. Do not allow the lysis reaction to proceed for more than 5 min.
3. Add 350 µl of Buffer N3 and invert the tube immediately but gently 4-6 times.
 To avoid localized precipitation, mix the solution gently but thoroughly, immediately after addition of Buffer N3. The solution should become cloudy.
4. Centrifuge for 10 min.
 A compact white pellet will form.
 During centrifugation, place a QIAprep spin column in a 2-ml collection tube.
5. Apply the supernatant from step 4 to the QIAprep column by decanting or pipetting.
6. Centrifuge 30-60 sec. Discard the flow-through.
7. [Optional]: Wash QIAprep spin column by adding 0.5 ml of Buffer PB and centrifuging 30-60 sec. Discard the flow-through.
 This step is necessary to remove trace nuclease activity when using endA⁻ strains such as the JM series, HB101 and its derivatives, or any wild-type strain, which have high levels of nuclease activity or high carbohydrate content. Host strains such as XL-1 Blue and DH5α[™] do not require this additional wash step.
8. Wash QIAprep spin column by adding 0.75 ml of Buffer PE and centrifuging 30-60 sec.

9. Discard the flow-through, and centrifuge for an additional 1 min to remove residual wash buffer.

! IMPORTANT: Residual wash buffer will not be completely removed unless the flow-through is discarded before this additional centrifugation. Residual ethanol from Buffer PE may inhibit subsequent enzymatic reactions.

10. Place QIAprep column in a clean 1.5-ml microfuge tube. To elute DNA, add 50 µl of Buffer EB (10 mM Tris-Cl, pH 8.5) or H₂O to the center of each QIAprep column, let stand for 1 min, and centrifuge for 1 min.

QIAprep Spin Miniprep Kit Protocol

using 5-ml collection tubes

The QIAprep Spin Miniprep procedure can be performed using 5-ml centrifuge tubes (e.g., Greiner, Cat. No. 115101 or 115261) as collection tubes to decrease handling. The standard protocol on pages 18-19 should be followed with the following modifications:

- Step 4: Place QIAprep spin column in a 5-ml centrifuge tube instead of a 2-ml collection tube.
- Step 6: Centrifuge at 3000 x g for 1 min using a suitable rotor (e.g., Beckman® GS-6KR centrifuge at ~4000 rpm). (The flow-through does not need to be discarded).
- Steps 7 & 8: For washing steps, centrifugation should be performed at 3000 x g for 1 min. (The flow-through does not need to be discarded).
- Step 9: Transfer QIAprep column to a microfuge tube. Microcentrifuge at maximum speed for 1 min. Continue with step 10 of the protocol.

- * Pellet cells at 10,000 rpm for 2 minutes
- * ~~Elute~~ Elute DNA in 50 µl's water instead of Buffer EB
- next time I do a miniprep
- * Store excess bacterial culture in 4°C

SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERSTOOD BY

Angela A. DePue

DATE

BAYER CORPORATION

RB55846

EXHIBIT

SUBJECT LBRI #147

tabbies

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Lisa Parkyn

03:35 PM

8

To: Janice Jackson/WESTH/PH/US/BAYER@BAYER-US-NOTES, Gwenda

9

cc: Ligon/WESTH/PH/US/BAYER@BAYER-US-NOTES
David Eustice/WESTH/PH/US/BAYER@BAYER-US-NOTES

10

Subject: LBRI Sequencing

11

Hi Jan and Gwen,

12

These are for the LBRI #147 program.

13

The vector is pCRII. The host was Top10 and the preps were done with the Qiagen kit.

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Thanks,

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Lisa

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1. RG_spi5_M13F
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3. RG_spi5_032
4. RG_spi5_033
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6. RG_spi5_047
7. RG_spi5_052
8. RG_spi5_M13R
9. RG_spi6_M13F
10. RG_spi6_029
11. RG_spi6_032
12. RG_spi6_033
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14. RG_spi6_047
15. RG_spi6_052
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17. RG_pla6_M13F
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25. RG_pla7_M13F
26. RG_pla7_029
27. RG_pla7_032
28. RG_pla7_033
29. RG_pla7_034
30. RG_pla7_047
31. RG_pla7_052
32. RG_pla7_M13R
33. RG_pla8_M13F
34. RG_pla8_029
35. RG_pla8_032

spi5, spi6, pla6, pla2

3x plasmid
2x primer
7x H₂O

pla7, pla8

1x plasmid
2x primer
9x H₂O

Primers

M13F, RG029, 032, 033,
034, 047, 052, M13R

36. RG_pla8_033
37. RG_pla8_034
38. RG_pla8_047
39. RG_pla8_052
40. RG_pla3_M13R
41. RG_pla3_M13F
42. RG_pla3_029
43. RG_pla3_032
44. RG_pla3_033
45. RG_pla3_034
46. RG_pla3_047
47. RG_pla3_052
48. RG_pla3_M13R

SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERST OD BY

Dargun A - DeHue

DATE

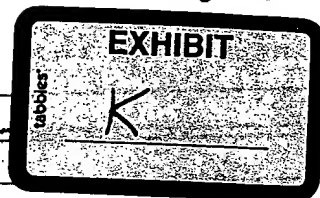
CROSS REFERENCES:

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RB55846

15

SUBJECT LBRI #147



① LifeTools Clustal W results comparing coolEST147_147spl5_2
the 147 consensus sequence (DNA sequence) (per vector)

Confidential - Property of Incyte Genomics, Inc. LifeTools Version 3.1 SeqServer

coolEST147_147spl5_2
coolEST147_147FLconsensus_2

CLUSTAL W (1.7) Multiple Sequence Alignments

Sequence format is Pearson
Sequence 1: coolEST147_147spl5_2 1767 bp
Sequence 2: coolEST147_147FLconsensus_2 1748 bp
Start of Pairwise alignments
Aligning...
Sequences (1:2) Aligned. Score: 100
Start of Multiple Alignment
There are 1 groups
Aligning...
Group 1: Sequences: 2 Score: 33212
Alignment Score 13534
CLUSTAL-Alignment file created (baaa03540.aln)
CLUSTAL W (1.7) multiple sequence alignment

```

coolEST147_147spl5_2      GAATTCGGCTCAGAGACCATGGAGAGGCACAGCCAGGGGAATGCATCTCC
coolEST147_147FLconsensus_2 -----CTCAGAGACCATGGAGAGGGACAGCCAGGGGAATGCATCTCC
*****

coolEST147_147spl5_2      AGCAAGAACACCTTCAGCTGGAGCATCTCCAGCCAGGCATCTCCAGCTG
coolEST147_147FLconsensus_2 AGCAAGAACACCTTCAGCTGGAGCATCTCCAGCCAGGCATCTCCAGCTG
*****

coolEST147_147spl5_2      GGACACCTCCAGGCCGGGCATCTCCAGCCAGGCATCTCCAGCCAGGCA
coolEST147_147FLconsensus_2 GGACACCTCCAGGCCGGGCATCTCCAGCCAGGCATCTCCAGCCAGGCA
*****

coolEST147_147spl5_2      TCTCCAGCTGGGACACCTCCGGGCCGGGCATCTCCAGCCAGGCATCTCC
coolEST147_147FLconsensus_2 TCTCCAGCTGGGACACCTCCGGGCCGGGCATCTCCAGCCAGGCATCTCC
*****

coolEST147_147spl5_2      AGCTGGTACACCTCCAGGCCGGGCATCTCCAGGCCGGGCATCTCCAGCCC
coolEST147_147FLconsensus_2 AGCTGGTACACCTCCAGGCCGGGCATCTCCAGGCCGGGCATCTCCAGCCC
*****

coolEST147_147spl5_2      AGGCATCTCCAGCCCGGGCATCTCCGGCTCTGGCATCACTTTCCAGGTCC
coolEST147_147FLconsensus_2 AGGCATCTCCAGCCCGGGCATCTCCGGCTCTGGCATCACTTTCCAGGTCC
*****

coolEST147_147spl5_2      TCATCCGGCAGGTGATCATCCGCCAGGTGAGCTCGGTGACAACTCCCC
coolEST147_147FLconsensus_2 TCATCCGGCAGGTGATCATCCGCCAGGTGAGCTCGGTGACAACTCCCC
*****

coolEST147_147spl5_2      AACCAGAGTGTACCTTGTAGAGCAACACCAAGTGGGGGCTGTACCCATCC
coolEST147_147FLconsensus_2 AACCAGAGTGTACCTTGTAGAGCAACACCAAGTGGGGGCTGTACCCATCC
*****

coolEST147_147spl5_2      GATCATCTCTCCAGGTGAGCAACCAAGTGGGGGCTGTACCCATCC
coolEST147_147FLconsensus_2 GATCATCTCTCCAGGTGAGCAACCAAGTGGGGGCTGTACCCATCC
*****
    
```

*Continued on
pg. 16

SIGNED BY Olga Parkyn

DATE

WITNESSED AND UNDERSTOOD BY Dr. Steven J. Diller

DATE

----- REFERENCES -----

SUBJECT LBRI #147

1	LifeTests Clustal W... (DNA Sequence)	
2		
3	coolEST147_147spl5_2	CCAGGTACGAGCCTGCCAAGTTCACCTGGCGGAGGGCCAGAAGCAGCT
4	coolEST147_147FLconsensus_2	CCAGGTACGAGCCTGCCAAGTTCACCTGGCGGAGGGCCAGAAGCAGCT
5	coolEST147_147spl5_2	ACCGCTCATCGGGTGGCTGCTCCTCCTCATTGCCCTGGTGGTTTCGCTCA
6	coolEST147_147FLconsensus_2	ACCGCTCATCGGGTGGCTGCTCCTCCTCATTGCCCTGGTGGTTTCGCTCA
7	coolEST147_147spl5_2	TCATCCTCTTCCAGTTCTGGCAGGGCCACACAGGGATCAGGTACAAGGAG
8	coolEST147_147FLconsensus_2	TCATCCTCTTCCAGTTCTGGCAGGGCCACACAGGGATCAGGTACAAGGAG
9	coolEST147_147spl5_2	CAGAGGGAGAGCTGTCCCAAGCACGCTGTTTCGCTGTGACGGGGTGGTGA
10	coolEST147_147FLconsensus_2	CAGAGGGAGAGCTGTCCCAAGCACGCTGTTTCGCTGTGACGGGGTGGTGA
11	coolEST147_147spl5_2	CTGCAAGCTGAAGAGTGACGAGCTGGGCTGGCTGAGGTTTGACTGGGACA
12	coolEST147_147FLconsensus_2	CTGCAAGCTGAAGAGTGACGAGCTGGGCTGGCTGAGGTTTGACTGGGACA
13	coolEST147_147spl5_2	AGTCTCTGCTTAAATCTACTCTGGGTCTCCCATCAGTGGCTTCCCATC
14	coolEST147_147FLconsensus_2	AGTCTCTGCTTAAATCTACTCTGGGTCTCCCATCAGTGGCTTCCCATC
15	coolEST147_147spl5_2	TGTAGCAGCAACTGGAATGACTCCTACTCAGAGAAGACCTGCCAGCAGCT
16	coolEST147_147FLconsensus_2	TGTAGCAGCAACTGGAATGACTCCTACTCAGAGAAGACCTGCCAGCAGCT
17	coolEST147_147spl5_2	GGGTTTCGAGAGTGCTCACCGGACAACCGAGCTTGCCCAAGGGATTG
18	coolEST147_147FLconsensus_2	GGGTTTCGAGAGTGCTCACCGGACAACCGAGCTTGCCCAAGGGATTG
19	coolEST147_147spl5_2	CCAACAGCTTCTCAATCTTGAGATACAATCCACCATCCAGGAAGCCTC
20	coolEST147_147FLconsensus_2	CCAACAGCTTCTCAATCTTGAGATACAATCCACCATCCAGGAAGCCTC
21	coolEST147_147spl5_2	CACAGGTCTGAATGCCCTTCCAGCGGTATATCTCCCTCCAGTGTCCCA
22	coolEST147_147FLconsensus_2	CACAGGTCTGAATGCCCTTCCAGCGGTATATCTCCCTCCAGTGTCCCA
23	coolEST147_147spl5_2	CTGCGGACTGAGGGCCATGACCGCGGATCTGGGAGGGCGCTGGCT
24	coolEST147_147FLconsensus_2	CTGCGGACTGAGGGCCATGACCGCGGATCTGGGAGGGCGCTGGCT
25	coolEST147_147spl5_2	CGGATAGCAAGTGGCTTGGCAAGTGAGTCTGCACTTGGCACCACCCAC
26	coolEST147_147FLconsensus_2	CGGATAGCAAGTGGCTTGGCAAGTGAGTCTGCACTTGGCACCACCCAC
27	coolEST147_147spl5_2	ATCTGTGGAGGCACGCTCATTGACGCCAGTGGGTGCTCACTGCCGCCA
28	coolEST147_147FLconsensus_2	ATCTGTGGAGGCACGCTCATTGACGCCAGTGGGTGCTCACTGCCGCCA
29	coolEST147_147spl5_2	CTGCTTCTTGTGACCCGGGAGAGGTCTGGAGGGCTGGAAGGTGTACG
30	coolEST147_147FLconsensus_2	CTGCTTCTTGTGACCCGGGAGAGGTCTGGAGGGCTGGAAGGTGTACG
31	coolEST147_147spl5_2	CGGGCACCAGCAACCTGCACCACTTGCCTGAGGCAGCTCCATTGCCGAG
32	coolEST147_147FLconsensus_2	CGGGCACCAGCAACCTGCACCACTTGCCTGAGGCAGCTCCATTGCCGAG
33	coolEST147_147spl5_2	ATCATCATCAACAGCAATTACACCGATGAGGAGGACGACTATGACATCGC
34	coolEST147_147FLconsensus_2	ATCATCATCAACAGCAATTACACCGATGAGGAGGACGACTATGACATCGC
	coolEST147_147spl5_2	CCTCATGCGGCTGTCCAAGCCCTGACCTGTCCGCTCACATCCACCTG
	coolEST147_147FLconsensus_2	CCTCATGCGGCTGTCCAAGCCCTGACCTGTCCGCTCACATCCACCTG
	coolEST147_147spl5_2	CTTGCCCTCCCATGCAATGGACAGACCTTTAGCCTCAATGAGACCTGCTGG
	coolEST147_147FLconsensus_2	CTTGCCCTCCCATGCAATGGACAGACCTTTAGCCTCAATGAGACCTGCTGG

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pg. 17.

SIGNED BY

Lisa Parkyn

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WITNESSED AND UNDERST OD BY

Dagmar A. Diller

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ADVICE RECEIVED.

BAYER CORPORATION

RB57846 17

SUBJECT LBRI #147

Lifetools Clustal w (DNA Sequence)

coolEST147_147sp15_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	3
coolEST147_147FLconsensus_2	ATCACAGGCTTTGGCAAGACCAGGAGACAGATGACAAGACATCCCCCTT	4
coolEST147_147sp15_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	5
coolEST147_147FLconsensus_2	CCTCCGGGAGGTGCAGGTCAATCTCATCGACTTCAAGAAATGCAATGACT	6
coolEST147_147sp15_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	7
coolEST147_147FLconsensus_2	ACTTGGTCTATGACAGTTACCTTACCCCAAGGATGATGTGTCTGGGGAC	8
coolEST147_147sp15_2	CTTCGTGGGGCCAGAGACTCCTGCCAGGAGACAGCGGGGGGCTCTTGT	9
coolEST147_147FLconsensus_2	CTTCGTGGGGCCAGAGACTCCTGCCAGGAGACAGCGGGGGGCTCTTGT	10
coolEST147_147sp15_2	CTGTGAGCAGAAACACCGCTGGTACCTGGCAGGTGTCAACAGCTGGGGCA	11
coolEST147_147FLconsensus_2	CTGTGAGCAGAAACACCGCTGGTACCTGGCAGGTGTCAACAGCTGGGGCA	12
coolEST147_147sp15_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACCAAAGTGACAGAA	13
coolEST147_147FLconsensus_2	CAGGCTGTGGCCAGAGAAACAAACCTGGTGTGTACACCAAAGTGACAGAA	14
coolEST147_147sp15_2	GTTCCTCCCTGGATTACACCAAGATGGAGAGCGAGGTCCGATTGAGAAA	15
coolEST147_147FLconsensus_2	GTTCCTCCCTGGATTACACCAAGATGGAGAGCGAGGTCCGATTGAGAAA	16
coolEST147_147sp15_2	ATCCTAACCCAGCTGGCCTGCTGCTCTGCACAGCACCAGGCTGCTGTGACTC	17
coolEST147_147FLconsensus_2	ATCCTAACCCAGCTGGCCTGCTGCTCTGCACAGCACCAGGCTGCTGTGACTC	18
coolEST147_147sp15_2	GAGAAAAAGCCGAATTC	19
coolEST147_147FLconsensus_2	GAGAAA	20

Protein Sequence

Sequence format is Pearson
 Sequence 1: coolEST147_147sp15+1_1 562 aa
 Sequence 2: coolEST147_147FLconsensus_3 562 aa
 Start of Pairwise alignments
 Aligning...
 Sequences (1:2) Aligned. Score: 100
 Start of Multiple Alignment
 There are 1 groups
 Aligning...
 Group 1: Sequences: 2 Score: 7726
 Alignment Score 3604
 CLUSTAL-Alignment file created [baaa03SM3.aln]
 CLUSTAL W (1.7) multiple sequence alignment

coolEST147_147sp15+1_1	MERDSHGNGASPARTPSAGASPAQASPAQASPAQASPAQASPAQASPAQAS	29
coolEST147_147FLconsensus_3	MERDSHGNGASPARTPSAGASPAQASPAQASPAQASPAQASPAQASPAQAS	30
coolEST147_147sp15+1_1	PGRASPAQASPAQASPAQASPAQASPAQASPAQASPAQASPAQASPAQAS	31
coolEST147_147FLconsensus_3	PGRASPAQASPAQASPAQASPAQASPAQASPAQASPAQASPAQASPAQAS	32
coolEST147_147sp15+1_1	SARSASVTTSPTRVYLVRATPVCAVPIRSSPARSAPATRATRESPTSLP	33
coolEST147_147FLconsensus_3	SARSASVTTSPTRVYLVRATPVCAVPIRSSPARSAPATRATRESPTSLP	34

*continued on
 pg. 18

SIGNED BY

Olga Paulsen

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WITNESSED AND UNDERSTOOD BY

Darguise A. DeHe

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EXHIBIT
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Sequence format is Pearson
Sequence 1: coolEST147_147spl5+1_1          562 aa
Sequence 2: coolEST147_147FLconsensus_3      562 aa
Start of Pairwise alignments
Aligning...
Sequences (1:2) Aligned. Score: 100
Start of Multiple Alignment
There are 1 groups
Aligning...
Group 1: Sequences: 2      Score: 7726
Alignment Score 3604
CLUSTAL-Alignment file created [baaa03SM3.aln]
CLUSTAL W (1.7) multiple sequence alignment

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coolEST147_147sp15+1_1      MERDSHGNGASPARTPSAGASPAQASPAGTTPGRASPAQASPAQASPAGTTP
coolEST147_147FLconsensus_3  MERDSHGNGASPARTPSAGASPAQASPAGTTPGRASPAQASPAQASPAGTTP
*****

coolEST147_147sp15+1_1      PGRASPAQASPAGTTPPGRASPGRASPAQASPARASPALASLSRSSSSGRSS
coolEST147_147FLconsensus_3 PGRASPAQASPAGTTPPGRASPGRASPAQASPARASPALASLSRSSSSGRSS
*****

coolEST147_147sp15+1_1      SARSASVTTSPTRVYLVLRATPVGAVPIRSSSPARSAPATRATRESPTGSLP
coolEST147_147FLconsensus_3 SARSASVTTSPTRVYLVLRATPVGAVPIRSSSPARSAPATRATRESPTGSLP

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*continued on
pg. 18

SIGNED BY

WITNESSED AND UNDERSTOOD BY

DATE**DATE** _____

SUBJECT LBRI #147

1	<u>Protein sequence (Life tools)</u>	
2		
3	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	KFTWREGQKQLPLIGCVLLIALVVSLLILFQFWQGHGTGIRYKEQRESCP KFTWREGQKQLPLIGCVLLIALVVSLLILFQFWQGHGTGIRYKEQRESCP *****
4		
5	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	KHAVRCDCGVVDCCKLKSDELQCVRFWDKSLKKIYSGSSHOWLPICSSNNW KHAVRCDCGVVDCCKLKSDELQCVRFWDKSLKKIYSGSSHOWLPICSSNNW *****
6		
7	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	DSYSEKTCQQLGFESAHRTEVAHRDFANSFSLRYNSTIQESLHRSECP DSYSEKTCQQLGFESAHRTEVAHRDFANSFSLRYNSTIQESLHRSECP *****
8		
9	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	SQRYISLQCSHCGLRAMTGRIVGGALASDSKWFQVSLHFGTTHICGGTL SQRYISLQCSHCGLRAMTGRIVGGALASDSKWFQVSLHFGTTHICGGTL *****
10		
11	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	IDAQWVLTAAHCFFVTREKVLGKWKVYAGTSLNLHQLPEAASIAEIIINSN IDAQWVLTAAHCFFVTREKVLGKWKVYAGTSLNLHQLPEAASIAEIIINSN *****
12		
13	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	YTDEEDDYDIALMRLSKPLTSLAHIHAPCLPMHGQTFSLNETCWTGFGK YTDEEDDYDIALMRLSKPLTSLAHIHAPCLPMHGQTFSLNETCWTGFGK *****
14		
15	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	TRETDDKTSPPFLREVQVNLIDFKKCNLYLVYDSYLTPRMCMAGDLGGGRD TRETDDKTSPPFLREVQVNLIDFKKCNLYLVYDSYLTPRMCMAGDLGGGRD *****
16		
17	coolEST147_147sp15+1_1 coolEST147_147FLconsensus_3	SCQGDGGGPLVCEQNNRWYLAGVTSWGTGCGQRNKPQVYTKVTEVLPWIIY SCQGDGGGPLVCEQNNRWYLAGVTSWGTGCGQRNKPQVYTKVTEVLPWIIY *****
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SIGNED BY

Lisa Parkyn

DATE

WITNESSED AND UNDERSTOOD BY

Dargines A. Adler

DATE

CROSS REFERENCES: